

Comparative Study of the Medicinal Values of *Indigoferatinctoria* and *Gossypium Hirsutum*

L. I. Ibrahim, A. Abdulazeez*, Z. M. Kolo, A. Usman, S. U. Nagenu, U. M. Badeggi, S. H. Mohammed., A. I. Muhammad and F. B. Salahuu

Received: 24 September 2021/Accepted 05 December 2021/Published online:28 December 2021

Abstract: Phytochemical screening was carried out using official and recommended analytical methods while the antimicrobial sensitivity test was implemented using the agar well diffusion method. From the phytochemical analysis of *indigoferatinctoria* extract, the results gotten, indicated the presence of significant concentrations of saponins, alkaloids and steroids, compared to the concentrations of flavonoids, tannins, glycoside, steroids (which were relatively minimal) and hydroxyl-anthraquinones (which was absent). The antimicrobial sensitive tests conducted on chloroform extract showed no activities against the selected bacteria, i.e *Salmonella typhi*, *E.coli*. Also, *Staphylococcus aureus* was not affected when treated with chloroform extract of *Indigoferatinctoria* but rather against ampicloxacillin control at 500 mg/ml The ethanolic extract showed minimal activities against *E. coli* and *Staphylococcus aureus* but not against *Salmonella typhi* Distilled water extract of *indigoferatinctoria* displayed activity against *E.coli* and *Staphylococcus aureus* but not against *Salmonella typhi*. Results recorded for phytochemical analysis indicated the presence of glycosides, alkaloids and steroids in all *Gossypium hirsutum* extract but not for tannins, flavanoid, and saponins extracts. The antimicrobial sensitivity test for antimicrobial of some selected organism against *gossypiumhirsutum* extract shows no activities when treated with the plant extract in both ethanol and distilled water extract but shows minimum activity with chloroform extract against *Salmonella typhi* at 80mg/ml and

100mg/ml. From the results obtained, *gossypiumhirsutum* extract showed no activity against the pathogen The antimicrobial activity of *Indigoferatinctoria* and *Gossypium hirsutum* may be harnessed for the treatment of infections caused by bacteria if toxicological tests confirm gives negative results

Keywords: *Indigoferatinctoria*, *Gossypium hirsutum*, phytochemical and analysis, antimicrobial.

L. I. Ibrahim

Department of Chemitry, Ibrahim Badamasi Babangida University, Lapai, Nigeria.

Email: <mailto:iilakan@ibbu.edu.ng>

Orcid id: 0000-0001-6764-4493

A. Abdulazeez*

Department of Chemitry, Ibrahim Badamasi Babangida University, Lapai, Nigeria

Email: abdulrahmanabdulazeez78@gmail.com

Orcid id: 0000-0001-8534-3536

Z. M. Kolo

Department of Chemitry, Ibrahim Badamasi Babangida University, Lapai, Nigeria.

A. Usman

Department of Chemitry, Nasarawa State University, Nasarawa, Nigeria.

Email: ausman2015@yahoo.com

Orcid id:0000-0003-0358-9984

S. U. Nagenu

Niger State College of Education, Minna, Nigeria

<https://journalcps.com/index.php/volumes>

Communication in Physical Science, 2021, 7(4): 370-377

U. M. Badeggi

Department of Chemistry, Ibrahim Badamasi Babangida University, Lapai, Nigeria.

Email: <mailto:umbadeggi@ibbu.ed.ng>

Orcid id: 0000-0003-4774-4237

S. H. Mohammed

Niger State College of Education, Minna, Nigeria

A. I. Muhammad

Department of Chemistry, Ibrahim Badamasi Babangida University, Lapai, Nigeria.

Email: <mailto:ibmaish@yahoo.com>

Orcid id: 0000-0002-4745-1453

F. B. Salahuu

Niger State College of Education, Minna, Nigeria

1.0 Introduction

A therapeutic plant is currently receiving more research interest than manufactured medications because of their mild or nontoxic nature (Seelananet al, 1997). The use of plants to cure or alleviate various medical ailments dates back to the crude days. Several investigations have confirmed that some plants characterized as wild have significant nutrient contents such as minerals, fundamental unsaturated fats and fiber substances (Tukan et al., 1998). Studies conducted by the world health organization revealed that between 65-80% of the population in developing nations relies on the customary prescription for their essential medicinal services and treatment. Because they believe that they are assessable, modest and are the product of known sources (Ashwini et al. 2011). *Indigoferatinctoria L* belongs to the family known as Fabaceae and some therapeutics benefits have been reported for the leaf of the plant, It is a sub bush; found in clammy deciduous timberlands and plain. The plant has hostile to lethal property (Warrireret al 2007). The flavonoid part of *Indigoferatinctorial* has been reported to exhibit some chemopreventive activity against benzeo pyrene-mediated lung

disease (Kameswaranet al.2008). Several studies have been reported for the chemical constituents of *Ingofera tinctorial L* and their impacts on its applications. Verma and Suresh (2002) published the phytochemical contents of leaf extract of *Indigoferatinctoria Linn* to include the presence of significant concentrations of alkaloids, terpenoids, glycosides and flavonoids. Adjerohet al., (2020) investigated the chemical composition of *Indigofera tinctorial pod* and reported moisture, crude fiber, carbohydrate, protein and ash contents to include 68.68, 14.83, 12.17, 2.63 and 1.01 % respectively. Phytochemical components identified from hot and cold ethanol extracts were alkaloids, flavonoids while tannins, saponins and cardiac glycosides were absent. Sharma and Agrawal (2015) investigated the potency of the aerial part of *Indigofera tinctorial L.* as a curative agent against some diseases using free radical and oxygen indices. The published results indicated that this plant part exhibit strong pharmaceutical activity and showed excellent antioxidant capabilities. Studies conducted by Srinivasanet al. (2015) towards the evaluation of the free radical scavenging potential using high performance thin layer chromatography gave results that indicated that the aqueous extract has mean concentrations of 267, 753.43, 349 mg/g of phenolics, flavonoids and antioxidants respectively. The extract also showed 2,2-diphenyl-1-picrylhydrazyl activity of 52.08 %, nitric oxide activity of 23.12 % and oxygen activity of 26.79 %).

Gossypium hirsutum L. is one of the most commonly encountered plants in our environment, exhibiting high yields. Comparative study of the medicinal values of *Indigoferatinctoria* and *Gossypium hirsutum*.(Kameswaran et al.2008).

2.0 Materials and Methods**2.1 Sample collection and preparation**

New *Indidoferatinctoria* and *Gossypium hirustum* were purchased randomly at PZ advertise from yankoli in Minna, Niger State.



The leaves were dried for constant weight at room temperature after which they grounded to powder form with mortar and pestle to expand the surface zone. The aqueous extract of the sample was prepared by soaking 100g of dried powder of each sample in 500cm³ of distilled water, chloroform and ethanol for 24 hours. The filtrate was placed on a water bath at 100°C for distilled water extract and 70°C for both chloroform and ethanol extract used for phytochemical screening and against antimicrobial screening.

2.2 Phytochemical screening

2.2.1 Preparation of reagents

10% sodium hydroxide solution was prepared by dissolving 10 g of sodium hydroxide pellet in 100 mL volumetric flask and made up to the mark with distilled water.

5% ferric chloride solution

was prepared by dissolving 5g of ferric chloride in 100 mL of the volumetric flask and was made up to the mark with distilled water.

50% H₂SO₄ solution

was prepared by diluting 50 mL of concentrated H₂SO₄ in a 100 mL volumetric flask and made up to the mark with distilled water.

10% HCl solution was prepared by diluting 10 mL of concentrated HCl in 100 mL of the volumetric flask and made up to the mark with distilled water.

13.35 g of Mercuric Chloride was dissolved in 60 mL of distilled water to give a solution A. 5g of Potassium Iodide was dissolved in 20 mL of distilled water to give a solution B. These two solutions (A and B) were mixed and made up to 1000 mL with distilled water in a 1000 mL volumetric flask (AOAC, 1990).

In the preparation of Wegner's reagent, 1.27g of a sublimed solution of iodine and 2 g of potassium iodide was dissolved in a beaker containing 20 mL of distilled water. This was transferred into a 1000 mL volumetric flask and made up to the mark with distilled water (AOAC, 1990).

2.2.2 Phytochemical screening of extract (Softwares 1993; Trease and Evans, 1989; Harborne, 1973).

To test for the presence of flavonoids, 3 cm³ aliquot of the filtrate was added to 1cm³ of 10% NaOH sodium hydroxide, if a yellow shading was built up this demonstrates the conceivable nearness of flavonoid compounds (El-Oleyiet al., 1994; Harbone, 1973).

The presence of tannins was carried out as follows: Ferric chloride solution 5% was added dropwise, to 2-3cm³ of the concentrate and the colour changed to blue-dark, demonstrating the nearness of tannins (Harbone, 1973; Trease and Evans, 1989).

To test for saponins, 10 cm³ of the concentrate was put in a test tube and blended in with 5cm³ of distilled water and shaken enthusiastically for stable relentless foam. The foaming was blended in with 3 drops of olive oil and shaken vivaciously, at that point watched for the formation of emulsion (Harbone, 1973).

The presence of glycosides was carried out by using Fehling solution (Harbone, 1973). The presence of alkaloids in the samples was confirmed using Wagner's and Mayer's reagents. The test for the presence of steroid was carried by the addition to the sample, in 2cm³ of chloroform, followed by the addition of 2 cm³ of tetraoxosulphate (VI) acid to obtain a dark reddish-brown dark colored at the interface (Harbone, 1973).

The steps employed to test for hydroxyl anthraquinones involve the shaking of 5 cm³ of the sample, followed by the addition of 10 cm³ benzene and 5 cm³ of 10% ammonia solution. The blend was shaken and the nearness of pink, red, or violet in the ammoniacal (lower) stage indicates the presence of anthraquinones (Harbone, 1973).

2.3 Determination of antibacterial assay

The affectability test was conducted using *Salmonella typhi*, *E.coli* and *Staphylococcus aureus* as references in the Agar well diffusion method. Nutrient Agar plates were set up and marked. A volume of 0.1 ml of each



institutionalized (McFarland standard) of the test life forms was vaccinated into independent plates utilizing spread plate strategy. A stopper borer was disinfected by dunking it into 70% liquor and going it through fire and is then used to drill four equidistant openings on the outside of the plate with one at the middle. One-tenth of the millimeter (0.1ml) of every one of the concentrated focus was brought into the four fringe openings while the gap in the inside would contain a control (Ampiclux). The agar plate was left for about an hour to permit the dispersion of the concentrates through the medium. They were then hatched at 37 °C for 24 hours. For each triplicate culture plate, the inhibition zone around each very much was then estimated and the mean worth was acquired and recorded in millimeter (mm).

2.3.1 Scoring and reading

The outcome was taken by considering the zone of development and inhibition of the living

organism by the test divisions. Activities and inactivity were observed in concurrence with the standard and adequate techniques (Hassan, 2004).

3.0 Results and Discussion

Table 1 presents results obtained from the phytochemical screening of leaf extract of *Indigoferatinctoria*. The results indicate that flavonoid and saponins were present in chloroform and aqueous extracts of *indigoferatinctoria* but absent in the ethanol extract. Saponins, alkaloids and steroids were present in all three extracts, namely, ethanol, chloroform and aqueous extracts. Glycosides and hydroxyl anthraquinones were absent in all the extracts. The results indicate that the aqueous extract contains many phytochemicals from *indigoferatinctoria* leaf, followed by chloroform extract and least by ethanol extract.

Table 1 Results of Phytochemical analysis of an extract from *Indigoferatinctoria*

Chemical constituents	Ethanol extract	Chloroform extract	Distilled water extract
Flavanoid	–	+	+
Tannins	–	–	+
Saponins	+	+	+
Glycosides	–	–	+
Alkaloids	+	+	+
Steroids	+	+	+
Hydroxyl-anthraquinones	–	–	–

From Table 2, which presents a list of phytochemicals detected in leaf extracts of *Gossypium hirsutum*, ethanol extract is observed to indicate the presence of all the phytochemicals except hydroxyl anthraquinones. The chloroform extract contained all except tannins, saponins and hydroxyl anthraquinones while the aqueous extract indicated the absence of flavonoids and hydroxyl anthraquinone only. Therefore the ethanol extract gave the highest concentration or yield of phytochemical while the chloroform extract presented the least. The antimicrobial sensitive tests results for chloroform extract of

Gossypium hirsutum leaf (Table 3) show no activities against the selected bacteria, that is, *Salmonella typhi*, *E.coli* and *Staphylococcus aureus* was not affected when treated with *Indigoferatinctoria* chloroform extract but rather against ampicloxacin control at 500 mg/ml. However, the ethanol extract of the leaf (Table 4) shows activities against *E. coli* and *Staphylococcus aureus* in minimum amount but there was no observable activity against *Salmonella typhi* while distilled water extract of *indigoferatinctoria* from also shows activity against *E.coli* and *Staphylococcus aureus* (Table 5 and not against *Salmonella typhi*).



Table 2 Results of Phytochemical analysis of *Gossypium hirsutum* leaves extract

Test	Distilled water	Ethanol	Chloroform
Flavonoid	+	+	+
Tannins	-	+	-
Saponins	+	+	+
Glycosides	+	-	+
Alkaloids	+	+	-
Steroids	+	+	+
Hydroxyl anthraquinones	-	-	-

Keys: - absent, + presence

Table 3 Chloroform extract of *Indigoferatinctoria* leaf

Organisms	Diameter of zone of inhibitor in "mm"			
	concentration in mg/ml			Ampicloxacillin 500 mg/ml
	60 mg/ml	80 mg/ml	100 mg/ml	
<i>Salmonella tyhi</i>	-	-	-	16.00
<i>E.coli</i>	-	-	-	10.00
<i>Staphylococcus aureus</i>	-	-	-	34.00

Table 4 Ethanolic extract of *Indigoferatinctoria* leave

Organism	Diameter of zone of inhibitor in "mm"			
	concentration in mg/ml			Ampicloxacillin 500 mg/ml
	60 mg/ml	80 mg/ml	100 mg/ml	
<i>Salmonella typhi</i>	-	-	-	10.00
<i>E.coli</i>	7.00	9.00	9.00	30.00
<i>Staphylococcus aureus</i>	6.00	8.00	9.50	35.00

Table .5: Distilled water extract of *indigoferatinctoria* leave extract

Organism	Diameter of zone of inhibitor in "mm"			
	concentration in mg/ml			Ampicloxacillin 500 mg/ml
	60 mg/ml	80 mg/ml	100 mg/ml	
<i>Salmonella typhi</i>	-	-	-	16.00
<i>E.coli</i>	5.00	8.00	11.00	30.00
<i>Staphylococcus aureus</i>	8.00	8.30	12.00	34.00



The sensitivity test for antimicrobial of some selected organism against *Gossypium hirsutum* leaf extract shows no activities when treated with the plant extract in both ethanol and distilled water extract but minimal activity was

observed for the chloroform extract against *Salmonella typhi* at 80 and 100 mg/ml respectively. Also, the *Gossypium hirsutum* leaf extract showed no activity against the pathogen.

Table 6 ethanol extract of *Gossypium hirsutum*

Organism	Diameter of zone of inhibitor in "mm"			Ampicloxacillin 500 mg/ml
	concentration in mg/ml			
	60 mg/ml	80 mg/ml	100 mg/ml	
<i>Salmonella typhi</i>	–	–	–	35.00
<i>E.coli</i>	–	–	–	25.00
<i>Staphylococcus aureus</i>	–	–	–	15.00

Table 7 Distilled water extract *Gossypium hirsutum*

Organism	Diameter of zone of inhibitor in "mm"			Ampicloxacillin 500 mg/ml
	concentration in mg/ml			
	60 mg/ml	80 mg/ml	100 mg/ml	
<i>Salmonella typhi</i>	–	–	–	10.00
<i>E.coli</i>	–	–	–	32.00
<i>Staphylococcus aureus</i>	–	–	–	32.00

Table 8 chloroform extracts *Gossypium hirsutum*

Organism	Diameter of zone of inhibitor in "mm"			Ampicloxacillin 500 mg/ml
	concentration in mg/ml			
	60 mg/ml	80 mg/ml	100 mg/ml	
<i>Salmonella Typhi</i>	–	3.00	3.20	12.00
<i>E.coli</i>	–	–	–	28.00
<i>Staphylococcus aureus</i>	–	–	–	30.00

4.0 Conclusion

Preliminary phytochemical evaluation of leaf extracts of *Indigoferatinctoria* and *Gossypium hirsutum* shows that the yield of phytochemical is strongly dependent on the choice of solvent. Ethanol seems to be the best solvent for *Gossypium hirsutum* but distilled water is the best solvent that gives the highest yield for phytochemicals in *Indigoferatinctoria*.

Hydroxyl anthraquinones were absent in all the extracts. Extracts from *Indigoferatinctoria* demonstrated displayed significant inhibition regarding the control. It is conclusive from the results of the study that synergistic interaction of the detected phytochemicals has some role in its biological activities.

5.0 References



- Adjero, L. A., Nwachukwu, M. O., Nnokwe, J. C., Azorji, J. N., Osinomumu, I. O. & Onyebuagu, P. C. (2020). Preliminary phytochemical screening and proximate analysis of *Indigoferatinctoria* L. (Uhe)Pod. *Amazonian Journal of Plant Research*, 4, pp. 3, pp. 639-645.
- Amrithpal, .(2006). *Medicinal plants of the world* Oxford & IBH".Publishing Co Pvt .Ltd New Delhi; 168.
- AOAC, (2002) *Association of Official Analytical analysis of International*, 17thEdition. William AOAC, Association of analytical chemists.
- Ashwini, .S., Gajalakshmi, S., Mythili, A. & Sathivelu, (2011). *Terminalia chebula*-A Pharmacological Review. *Journal of Pharmacy Research*, 4, 9, pp. 2884-2887.
- Bakasso, S., Lamien, M., Lamien, C. E., Keindrebeogo, M., Millogo, J., Ouedraogo A. G. & Nacoulma, A.G. (2008). Polyphenol contents and antioxidant activities of five *Indigoferaspecies* (Fabaceae) from Burkina Faso, *Pakistan Journal of Biological Sciences*, .11, pp. 1435-1442.
- Bisignano, G., Germano, M. P., Nostro, A. & Sanogo, R. (1996). Drugs used in Africa as dyes: antimicrobial activities. *Phytotherapy Research*, 9: pp. 346-350.
- Carter, C .L. & Chesney, M. C. (1949). Hiptagenic acid identified as beta-nitropropionic acid, *Nature*, 164, pp. 575-576.
- Culvenor, C.C., Foster, M. C. & Hegarty, M. P. (1971). A total synthesis of indospicine, 6-Amidino-2-aminohexanoic acid,. *Australian Journal of Chemistry*, .24, pp. 371-375.
- Eisenbrand G., Hippe F., Jakobs S. & Muehlbeyer S. (2004). Molecular mechanisms of indirubin and its derivatives: novel anticancer molecules with their origin in traditional Chinese phytomedicine. *Journal of Cancer Research and Clinical Oncology*, 130, pp. 627-635.
- Hartwell, J. L. (1967–1971). *Plants used against cancer*. A survey. *Lloydia* , pp. 30–34.
- Kameswaran R & Ramanibhai R. (2008). Protective effect of flavonoids fraction of *Indigoferatinctoria* on Benzopyrene induced Lung carcinogenicity in Swiss Albino mice. *International Journal of Cancer Research*, 4, pp. 71-80.
- List, P.H. & Horhammer, L. 1969–1979. Hager's handbuch der pharmazeutischen praxis. vols 2–6. Springer-Verlag, Berlin.
- Martinez, C., Calero, A., Dominguez ,X. A & Hinojosa, M.(1978). Louisfieserone, an unusual flavanone from *Indigoferasuffruticosa*, *Tetrahedron Letters*.5, pp. 429-432.
- Mujeeb, F., Bajpai, P., & Pathak, N. (2014). Phytochemical Evaluation, Antimicrobial activity, and determination of bioactive components from leaves of *Aeglemarmelos*. *BioMedica Research*, <http://doi.org/10.1-155/2014/497606>.
- Muthulingam, M. Mohandoss, P. Indra, N. & Sethupathy, S.(2010).Antihepatotoxic efficacy of *Indigoferatinctoria* (Linn.) on paracetamol induced liver damage in rats. *IJPBR*;1(1):13-18
- Nadkarni KM. (2005). *Indian Plants and Drugs*. New Delhi: Srishti Book Distributors; pp.172-3.
- Narender, T., Khaliq, T., Puri ,A. & Chander, R. (2006). Antidyslipidemic activity of furano-flavonoids isolated from *Indigoferatinctoria*, *Bioinorganic and Medicinal Chemistry*, .16, pp. 3411-3414.
- Seelanan, T., Schnabel A & Wendel, J. F. (1997). Congruence and consensus in the cotton tribe (Malvaceae). *Syst Bot.*;22, 2, pp. 259–90.
- Sharma, V. & Agarwal, A. (2015). Phytochemical and antioxidant assays of methanol and hydromethanol extracts of ariel parts of *Indigofera tinctorial* Linn. *Indian Journal of Pharmaceutical Science*, 77, 6, pp. 729-734.
- Sheikh, N. Kumar, Y. Misra, A. K. and Pfoz, L. 2013. Phytochemical screening to validate the ethnobotanical importance of root tubers of *Dioscorea* species of Meghalaya, North



- East India. *Journal of Medicinal Plants Studies*, 1, pp. 62–69.
- Sharma, V. & Agarwal, A. (2015). Phytochemical and antioxidant assays of methanol and hydromethanol extracts of aerial parts of *Indigofera tinctoria* Linn. *Indian Journal of Pharmaceutical Science*, 77, 6, pp. 729-734.
- Srinivasan, S., Wankhar, W., Rathinasamy, S. & Rajan, R. (2015). Free radical scavenging potential and HPTLC analysis of *Indigoferatinctorialinn* (Fabaceae), *Journal of Pharmaceutical Analysis*, 6, 2, pp. 125-131.
<https://doi.org/10.1016/j.jpha.2015.04.003>.
- Subramaniam, S. and Ayarivan, P. (2014). Cytokine mediated immunomodulatory properties of kaempferol-5-O- β -D-glucopyranoside from methanol extract of aerial parts of *Indigofera aspalathoides* Vahl ex DC Swarnalatha, Ayarivan Puratchikody *International Journal of Research in Pharmaceutical Sciences*, 5, pp. 1-6.
- Tukan, S. K., Takruri, H. R. and Al-Eisaw, D. M. (1998). The use of wild edible plants in the Jordanian diet. *International Journal of Food Sciences and Nutrition* 49, pp.225 - 235.
- Verma S M, Suresh KB and Verma Amit(2010). Antidiabetic Activity of Leaves *Indigoferatinctoria* Linn (Fabaceae) *International Journal of Toxicological and Pharmacological Research.*:1, 2, pp. 42-43.
- Verma, S. M. & Suresh, K. B. (2002). Phytochemical investigations of *indigoferatinctorialinn* leaves. *Anc Sci Life.*, 21, 4, pp. 235-239
- WarrierPK, NambiarVPK and C Ramankatty(2010). *Indian medicinal plants*. Published by Orient Longman Private Limited. Chennai, 3, pp. :210-213
- Warrirer . P. K., Nambiar, V. P. K. & Ramankatty, C. (2007). *Indian medicinal plants*. Chennai: Orient Longman Private Limited, pp. 210-213.
- Wendel, J. F., Brubaker, C. L. & Seelanan, T. (2010). *The origin and evolution of Gossypium*. In: Stewart, J. M. et al., editors, *Physiology of cotton*. The Netherlands: Springer; pp.. 1–18.
- Conflict of Interest**
 The authors declared no conflict of interest

